

AD-A221 535

DTIC FILE COPY



Effects of Ultraviolet Radiation on the Oxygen Uptake Rate of the Rabbit Cornea

(Reprint)

By

Morris R. Lattimore

Sensory Research Division

July 1989



Approved for public release; distribution unlimited.

90 05 01 010

**United States Army Aeromedical Research Laboratory
Fort Rucker, Alabama 36362-5292**

Notice

Qualified requesters

Qualified requesters may obtain copies from the Defense Technical Information Center (DTIC), Cameron Station, Alexandria, Virginia 22314. Orders will be expedited if placed through the librarian or other person designated to request documents from DTIC.

Change of address

Organizations receiving reports from the U.S. Army Aeromedical Research Laboratory on automatic mailing lists should confirm correct address when corresponding about laboratory reports.

Animal use

In conducting the research described in this report, the investigators adhered to the Guide for care and use of laboratory animals, as promulgated by the Committee on Care and Use of Laboratory Animals of the Institute of Laboratory Resources Commission on Life Sciences, National Academy of Sciences-National Research Council.

Disposition

Destroy this report when it is no longer needed. Do not return it to the originator.

Disclaimer

The views, opinions, and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy, or decision, unless so designated by other official documentation. Citation of trade names in this report does not constitute an official Department of the Army endorsement or approval of the use of such commercial items.

Reviewed:



BRUCE C. LEIBRECHT, Ph.D.
LTC, MS
Director, Sensory Research
Division



J. D. LaMOTHE, Ph.D.
COL, MS
Chairman, Scientific
Review Committee

Released for publication:



DAVID H. KARNEY
Colonel, MC
Commanding

Unclassified

SECURITY CLASSIFICATION OF THIS PAGE

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
1a. REPORT SECURITY CLASSIFICATION Unclassified			1b. RESTRICTIVE MARKINGS		
2a. SECURITY CLASSIFICATION AUTHORITY			3. DISTRIBUTION / AVAILABILITY OF REPORT Approved for public release, distribution unlimited		
2b. DECLASSIFICATION / DOWNGRADING SCHEDULE					
4. PERFORMING ORGANIZATION REPORT NUMBER(S) USAARL 89-11			5. MONITORING ORGANIZATION REPORT NUMBER(S)		
6a. NAME OF PERFORMING ORGANIZATION U.S. Army Aeromedical Research Laboratory		6b. OFFICE SYMBOL (If applicable) SGRD-UAS-VS	7a. NAME OF MONITORING ORGANIZATION U.S. Army Medical Research and Development Command		
6c. ADDRESS (City, State, and ZIP Code) Fort Rucker, AL 36362-5292			7b. ADDRESS (City, State, and ZIP Code) Fort Detrick Frederick, MD 21701-5012		
8a. NAME OF FUNDING / SPONSORING ORGANIZATION		8b. OFFICE SYMBOL (If applicable)	9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER		
8c. ADDRESS (City, State, and ZIP Code)			10. SOURCE OF FUNDING NUMBERS		
			PROGRAM ELEMENT NO.	PROJECT NO.	TASK NO.
			WORK UNIT ACCESSION NO.		
11. TITLE (Include Security Classification) Effects of Ultraviolet Radiation on the Oxygen Uptake Rate of the Rabbit Cornea					
12. PERSONAL AUTHOR(S) Lattimore, Morris R., Jr.					
13a. TYPE OF REPORT		13b. TIME COVERED FROM _____ TO _____		14. DATE OF REPORT (Year, Month, Day) 1989 July	
				15. PAGE COUNT 6	
16. SUPPLEMENTARY NOTATION This work was done at the University of Houston and this report is a reprint of a publication in Optometry and Vision Science, Volume 66, Number 2, 1989					
17. COSATI CODES			18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)		
FIELD	GROUP	SUB-GROUP			
06	04				
06	05		cornea, metabolic activity, oxygen uptake rate, protection, ultraviolet radiation		
19. ABSTRACT (Continue on reverse if necessary and identify by block number) Ultraviolet radiation (UVR) has been demonstrated to be involved in a number of adverse ocular effects. One aspect of UVR-induced corneal stress only recently documented is an alteration of epithelial energy metabolite levels. In this study, in order to examine wavelength and dose dependency issues concerning metabolic effects of UVR, exposures were made at four different wavelengths (290, 300, 310 and 360 nm) and five different mean radiant exposures (0.05, 0.10, 0.15, 0.20 and 0.25 J-cm ⁻²). Pre- and postexposure levels of relative metabolic activity of the corneal epithelium were monitored <i>in vivo</i> by recording the corneal oxygen uptake rate with a micropolarographic electrode. A paired difference analysis demonstrated a decrease in relative corneal metabolic activity that was both wavelength- and dose-dependent. These relative metabolic effects provide some insight toward the understanding of underlying damage mechanisms, and imply a broader radiant energy susceptibility range of the eye than previously thought.					
20. DISTRIBUTION / AVAILABILITY OF ABSTRACT <input checked="" type="checkbox"/> UNCLASSIFIED/UNLIMITED <input type="checkbox"/> SAME AS RPT. <input type="checkbox"/> DTIC USERS			21. ABSTRACT SECURITY CLASSIFICATION Unclassified		
22a. NAME OF RESPONSIBLE INDIVIDUAL Chief, Scientific Information Center			22b. TELEPHONE (Include Area Code) (205) 255-6907		22c. OFFICE SYMBOL SGRD-UAX-SI

Effects of Ultraviolet Radiation on the Oxygen Uptake Rate of the Rabbit Cornea

MORRIS R. LATTIMORE, JR.*

Major, Medical Service Corps, U.S. Army, United States Army Aeromedical Research Laboratory, Fort Rucker, Alabama

ABSTRACT

Ultraviolet radiation (UVR) has been demonstrated to be involved in a number of adverse ocular effects. One aspect of UVR-induced corneal stress only recently documented is an alteration of epithelial energy metabolite levels. In this study, in order to examine wavelength and dose dependency issues concerning metabolic effects of UVR, exposures were made at four different wavelengths (290, 300, 310 and 360 nm) and five different mean radiant exposures (0.05, 0.10, 0.15, 0.20, and 0.25 J·cm⁻²). Pre- and postexposure levels of relative metabolic activity of the corneal epithelium were monitored *in vivo* by recording the corneal oxygen uptake rate with a micropolarographic electrode. A paired difference analysis demonstrated a decrease in relative corneal metabolic activity that was both wavelength- and dose-dependent. These relative metabolic effects provide some insight toward the understanding of underlying damage mechanisms, and imply a broader radiant energy susceptibility range of the eye than previously thought.

Key Words: cornea, metabolic activity, oxygen uptake rate, protection, ultraviolet radiation

The electromagnetic spectrum has been divided into a number of discrete regions based on wavelength. The regions labeled as UV, visible, and infrared are of immediate interest for those concerned about ocular effects of nonionizing radiant energy. Shorter wavelength radiation has a higher potential energy¹ and, therefore, a greater capacity for tissue damage, if absorbed. As a result, the UV region has gained more investigative attention than the others. The UV region has been subdivided into several bands based on apparent biologic effect: UV-C, 200 to 290 nm; UV-B, 290 to 320 nm; and UV-A, 320 to 400 nm. Although these divisions are not agreed upon absolutely because of some phenomenological variation, generally the terminology

has been accepted. Research techniques have become increasingly sophisticated, allowing the detection of subtle functional changes that can occur in response to corneal stress or insult. These methodologies have introduced the possibility of detecting changes in corneal function as a result of UV radiant exposures that may not necessarily result in histologically detectable damage.

The action spectrum for histologically detectable corneal damage from exposure to UVR has been said to begin at 210 nm and extend to 315 nm.²⁻⁷ Previously the corneal radiant exposure data above 320 nm have not been considered part of the action spectrum of the cornea because the exposure levels necessary to produce minimal histologic damage to the cornea are comparably high.⁸

However, recent data demonstrate an alteration of corneal epithelial energy metabolites in the pigmented rabbit exposed *in vivo* to UVR up to 360 nm at radiant exposures that exceeded, were at, or were below histologic damage threshold values.⁹ This finding suggests the need for increased investigative attention toward the effects of UVR. Most discussions of UVR center on histologic findings and cell death; the study of previously undetectable functional changes may reveal information concerning the underlying damage mechanism(s) of UVR, and shed light on possible recovery processes.

The four experimental wavelengths (290, 300, 310, and 360 nm) were chosen based on an interest in maintaining an environmental relevance, inasmuch as 290 nm UVR and above can be found at the earth's surface.¹⁰ An additional factor was the intention of creating a distinctive span of effects because the corneal thresholds for histologic damage (H_c) vary considerably. The corneal radiant exposure H_c in the rabbit ranges from 0.012 J·cm⁻² at 290 nm to 0.022 J·cm⁻² at 300 nm to 0.05 J·cm⁻² at 310 nm to near 65 J·cm⁻² at 360 nm. By varying both the wavelength and the radiant exposure, it was predicted that effects on the oxygen uptake rate might vary from severe at 290 nm to moderate at 300 and 310 nm to nonexistent at 360 nm. Lastly, the source output happens to peak in these regions, thereby helping minimize some of the time differences typical of a noncoherent source exposure.

Presented at the Annual Meeting of the American Academy of Optometry, Denver, Colorado, December, 1987.

Received July 18, 1988; revision received October 12, 1988.

* Optometrist, Ph.D.

UV Effects on Corneal Oxygen Uptake—Lattimore 117

Availability Codes

Dist Avail and/or
Special

A-1 20

METHODS

Exposure Instrumentation

Source calibration and radiometric quantification duplicated the procedures described by Pitts et al.⁸ The UVR source was a 5000 W xenon-mercury (Xe-Hg) high pressure arc lamp, powered by a 10 kW direct current power supply regulated to $\pm 0.5\%$, and capable of delivering from 0 to 80 amps at 25 to 65 V to the arc electrodes. The lamp housing was cooled by a double blower system. The radiation from the source was focused at a double monochromator entrance slit by the housing optics. A 10 cm quartz-enclosed water chamber was placed between the focusing lenses and the monochromator to remove the infrared radiation. The exit optical beam was focused by a quartz lens with a beam size of 1.6 by 1.8 cm at the plane of the experimental animal's cornea.

The desired UVR waveband was obtained with a Czerny-Turner double grating monochromator (model 25-100; Jarrell Ash Division, Fisher Scientific, Waltham, MA) possessing a double mirror setup with gratings blazed at 300 nm and grooved with 1180 grooves per nm, allowing approximately a 5.0-nm bandpass. The linear dispersion equates to a value of 0.82 nm per millimeter. Entrance, intermediate, and exit slits were set to pass a nominal full bandpass of 6.6 nm. The double monochromator system was aligned with a helium-neon laser and the wavelength counter was calibrated with a mercury source.

Exposure durations were set with a Gerbands electronic shutter (Ralph Gerbands Co. Inc., Arlington, MA) controlled by a Hewlett-Packard model 5330B preset counter. The present counter allowed exposure durations of any desired length with millisecond accuracy.

Source Measurement

An Eppley 16 junction thermopile (Eppley Laboratory Inc., Salem, MA), traceable to a National Bureau of Standards standard source, was used to characterize the spectral irradiance of the UVR source. When taking the spectral irradiance readings, the thermopile was placed in the same position relative to the monochromator exit port as the rabbit's cornea was to be situated during UVR exposure. The irradiance (E_e), in watts per square centimeter ($\text{W}\cdot\text{cm}^{-2}$), incident on the thermopile was determined by the following relation:

$$E_e = kV_t$$

The value "k" represents the thermopile calibration constant in microwatts per square centimeter per microvolt ($\mu\text{W}\cdot\text{cm}^{-2}\cdot\mu\text{V}^{-1}$), whereas the value " V_t " represents the thermopile-voltmeter reading in microvolts (μV). The calibration constant for the thermopile used in this experiment was $5.131 \mu\text{W}\cdot\text{cm}^{-2}\cdot\mu\text{V}^{-1}$. The radiant exposure (H), in Joules per square centimeter, was calculated by the formula H

$= E_e t$. The value "t" is simply time in seconds; it should be kept in mind that a Joule is a watt-second. Therefore, for a given irradiance (E_e), the exposure duration (t) can be varied to obtain different values of radiant exposure (H) as desired. The above means of output characterization and source calibration was estimated to have a $\pm 10\%$ accuracy.

The variation of t in order to obtain a constant "H," in the context of the wavelengths used in this experiment, creates an outcome that is somewhat dependent upon the validity of the principle of reciprocity (i.e., the biologic effects or endpoints are independent of exposure time and irradiance). Corneal effects of a krypton-ion laser, with simultaneous output at 350.7 and 356.4 nm (3:1 ratio), illustrates that the product of threshold intensity and the pulsewidth is a constant; the thresholds for multi-pulse exposures have been shown to be in agreement with those for single-pulse exposures.¹¹ A similar corneal damage pattern can be elicited from helium-cadmium laser data at 325 nm.¹² Based on the literature, it is reasonable to assume that reciprocity holds for all four UVR wavelengths used in this experiment.

Experimental Animals

Healthy, adult, pigmented New Zealand rabbits were used as the experimental animals. All animals were procured from a single source to ensure constant breeding practices. The animals were housed in NIH-approved quarters under normal, controlled (12 h on, 12 h off) lighting conditions. The animals were maintained, and the experiments were conducted in accordance with procedures outlined in the "Guide for Laboratory Animal Facilities and Care" of the National Research Council, National Academy of Sciences. Anesthesia was maintained throughout the course of the experiment with intramuscular injections of Ketalar (ketamine hydrochloride) (10 mg/kg) and Rompun (5 mg/kg).

Before exposure, each eye was examined with a biomicroscope; animals with anomalies of the cornea were rejected. The animals were restrained in a specially designed holder with only one eye per animal being exposed. The cornea was centered normal to the monochromator exit beam while the monochromator was set in the visible range. The eye then was exposed to UVR at one of the four experimental wavelengths (290, 300, 310, or 360 nm) for specific, predetermined radiant exposure durations. All exposure sessions took place at the same time of day for each experimental group, with ambient illuminance kept constant for all O_2 uptake studies.

Oxygen Electrode

The micropolarographic oxygen probe consisted of a platinum cathode (25 μm diameter) and a silver anode embedded in a plastic carrier. A potassium chloride (KCl) solution served as an electrolytic

bridge between the cathode and the anode. An oxygen-permeable polyethylene membrane, 25 μm thick, effectively sealed the entire electrode-KCl assembly into one operating unit. The micropolarographic system was similar to that used by Benjamin and Hill.¹³

The experimental procedure involved applying the probe to the anterior surface of the corneal epithelium of the living anesthetized rabbit. The sensor, when applied to the eye, provided a limited reservoir of oxygen for the underlying tissue. The average rate of oxygen depletion from the sensor reservoir, between recordings of 140 mm Hg and 40 mm Hg, and after correction for the system-specific time constant, became the measure of the corneal oxygen uptake rate. This, in turn, represents only a relative measure of the aerobic requirement of the cornea, because the extent that the epithelium, stroma, and endothelium each contribute to the corneal oxygen uptake rate has not been adequately established. Published estimates for the epithelium range from 55%¹⁴ to 70%,¹⁵ with an unpublished estimate ranging as high as 93% (WJ Benjamin and M Zagrod, personal communication, 1988).

Micropolarographic Application

The eyes of 16 rabbits were exposed *in vivo* to specific exposure levels of UVR. There were four rabbits in each experimental group. Before UVR exposure, five baseline oxygen uptake recordings were made for each eye in the fashion previously described. Oxygen uptake recordings were made again, 2 min after UVR exposure was discontinued, enabling the experimenter to compare the change in oxygen uptake rate resulting from the UVR exposure. The uptake rate values were subjected to a paired-difference analysis. Because the postexposure reading was a "one-time" reading, the standard error of the mean baseline reading was used as an estimate of the postexposure error. The unexposed eyes were also monitored pre- and postexposure to assess the stability of the readings. A two-way analysis of variance was performed in order to estimate differences both within and among experimental groups, and to determine the presence or absence of an overall effect of UVR on the measured oxygen uptake rates.

RESULTS

The radiant exposure for all experimental wavelengths was varied from 0.10 to 0.25 $\text{J}\cdot\text{cm}^{-2}$ in 0.05 $\text{J}\cdot\text{cm}^{-2}$ steps. The mean exposure levels were selected from a larger data-set that examined dose issues. The corneal oxygen uptake rate was measured 2 min after the UVR exposure was discontinued. A difference comparison between the pre-exposure baseline and the postexposure oxygen uptake rate demonstrated a wavelength- and dose-specific effect; see Table 1 for a summary data chart.

By plotting the UVR-altered corneal oxygen up-

TABLE 1. Corneal oxygen uptake rate data.*

Wavelength (nm)	Radiant Exposure ($\text{J}\cdot\text{cm}^{-2}$)	Decreased Corneal Oxygen Uptake Rate (mm Hg $\text{O}_2\cdot\text{sec}^{-1}$)	SE (of Baseline)
290	0.10	-2.67	0.22
290	0.15	-3.47	0.21
290	0.20	-4.17	0.21
290	0.25	-4.81	0.23
300	0.10	-1.08	0.22
300	0.15	-1.43	0.20
300	0.20	-1.75	0.21
300	0.25	-2.05	0.21
310	0.10	-0.57	0.21
310	0.15	-0.76	0.20
310	0.20	-0.95	0.21
310	0.25	-1.12	0.23
360	0.10	-0.10	0.23
360	0.15	-0.16	0.20
360	0.20	-0.21	0.22
360	0.25	-0.26	0.22

* Summary data chart outlining the four wavelengths and four radiant exposures that were used, plus the resultant paired decrease in the oxygen uptake rate. The SE is from the pre-exposure baseline data and is reported in an attempt to portray at least an estimate of the experimental error. This same comment applies to the error bars seen in both Fig. 1 and Fig. 2.

take rate data as a function of wavelength, and by making separate data-sets for each radiant exposure, a "family" of plots is obtained (Fig. 1). A two-way analysis of variance demonstrated an overall significant between-groups difference ($p < 0.0001$), as well as revealed an interactive effect between wavelength and dose ($p < 0.005$). Unexposed eyes exhibited no significant change in corneal oxygen uptake rates over the course of the experiment.

DISCUSSION

Fig. 1 presents the decrease in corneal oxygen uptake rate as a function of wavelength with separate plots for each level of radiant exposure. The alteration in corneal oxygen uptake is both wavelength- and dose-dependent, suggesting the presence of at least two different mechanisms of action, based on the differentiation of the three regions of the UV spectrum as discussed in the introduction. The importance here is that different proposed damage mechanisms, unique to the different regions of the UV spectrum, appear to have the common effect of decreasing the corneal oxygen uptake rate and, presumably, metabolic activity. Speculation regarding the possible mechanism(s) responsible for this alteration in oxygen uptake would have to take into account the fact that the change was registered 2 min after the exposure was discontinued.

Current damage mechanism theories involve DNA structural alteration,^{16,18} generalized changes in enzymatic activity,¹⁹⁻²³ and/or changes in mitochondrial activity.²⁴⁻²⁶ Because the metabolic alteration suggested by the decrease in the corneal oxygen uptake rate is essentially immediate, it would

Wavelength- and Dose-Dependency Effects of UVR on the Corneal Oxygen Uptake Rate

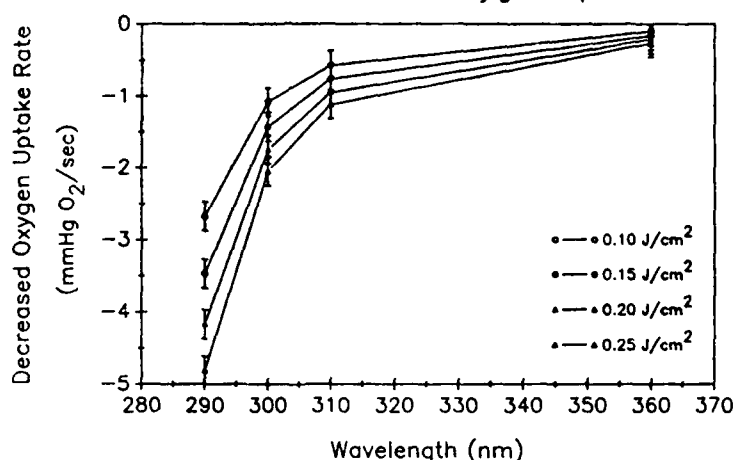


Figure 1. The 16 data points depict four "families" of curves ascending toward normal levels of corneal oxygen uptake as the UVR dose decreases and as the UVR wavelength increases. The error bars represent the SE of the pre-exposure baseline mean before the difference analysis.

be unlikely that DNA structural alteration is responsible. A change in overall enzymatic activity, or a change in mitochondrial activity, or both mechanisms acting "in concert" could be responsible for this UVR-induced decrease in the measured corneal oxygen uptake rate.

The demonstrated UVR-alteration of the corneal oxygen uptake rate implies an alteration in the relative metabolic activity that is graded in nature, suggesting the absence of a true threshold effect. Therefore, even a minimal dose could have some detrimental effect on corneal tissue function. When an already metabolically stressed cornea is exposed to a radiation source for a prolonged period of time, the adverse metabolic effect may be compounded. This has potential clinical significance when applied to contact lens wearers; extended periods of outdoor activity could be subjecting the cornea to a doubly stressful situation: decreased oxygen availability as a result of contact lens wear, and decreased oxygen utilization as a result of the sunlight's radiant energy. Although the conclusion that UVR is capable of causing a decrease in metabolic activity is specific to the cornea, it might be more generally extended to other tissues.

UV-A has been clearly linked with cataract development associated with certain phototoxic pharmaceutical compounds.²⁹⁻³³ Evidence also has been presented for the noninteractive induction of cataracts by both UV-B³⁴ and UV-A.³⁵ Although epidemiologic studies have sought to establish a possible relation between sunlight exposure and cataract prevalence,³⁶⁻⁴² some investigators have suggested the presence of a specific link between sunlight exposure and individual cataract development.^{35, 43} The retina, as well, has been shown to be

subject to damage as a result of exposure to radiant energy; by UV-B exposure,^{44, 46} as well as by UV-A exposure.⁴⁷

In fact, some of these experimental UVR exposures have induced cataracts or caused retinal damage at levels close to those available from midday solar exposure.⁴⁸ In addition, visible radiation has been implicated in retinal damage processes.⁴⁹⁻⁵⁸ Further research along this line might provide significant steps toward understanding the mechanism of blue light damage to the retina.^{49, 51, 53, 56}

The fact that computerized extrapolations of the data plots illustrated in Fig. 1 intersect the pre-exposure, normalized baseline at a mean wavelength of 447 nm (Fig. 2) suggests that the cornea could be vulnerable to a decrease in metabolic activity when exposed to radiation extending into the visible spectrum. The results of this study highlight the issue of adequate visual protection. At one time 320 nm was considered to be an adequate protection cut-off value for the entire eye.⁶ Later studies revealed 340 to 360 nm radiation to be potentially harmful to the eye in general.^{7, 59, 60} Now it is intimated that short wavelength, visible radiation might be stressful to the cornea, and perhaps similarly affect other ocular tissues. In conclusion, all UVR sources (including some recently publicized diagnostic systems),⁶¹⁻⁶⁴ and perhaps visible short wavelength sources as well, regardless of the energy output or the waveband, should be regarded with greater caution and used in a more prudent fashion.

ACKNOWLEDGMENTS

I thank William J. Benjamin, Jr., O.D., Ph.D. and Donald G. Pitts, O.D., Ph.D.

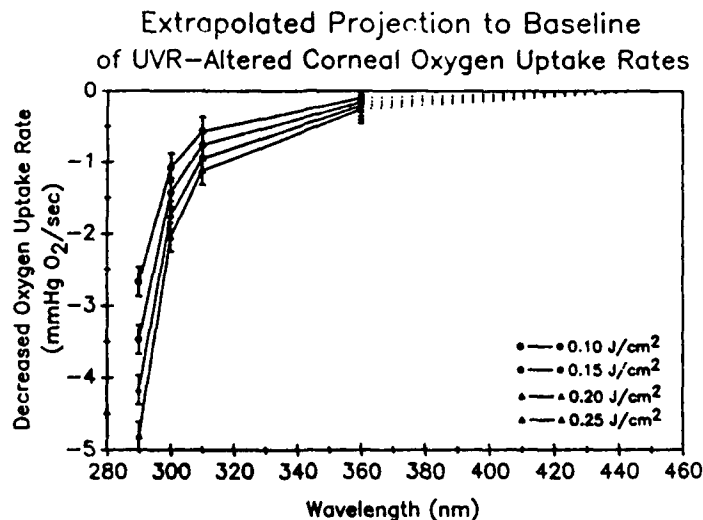


Figure 2. An extrapolated regression analysis of the four UVR wavelength groups reveals a mean return to baseline at 447 nm, suggesting a metabolic efficacy of short wavelength, visible radiation. Again, the error bars represent the SE of the pre-exposure baseline mean before the difference analysis.

REFERENCES

- Planck M. Dtsch Phys Ges 1900:237. In: Duke-Elder S, ed. *System of Ophthalmology*, vol 4. The Physiology of the Eye and of Vision. St Louis: CV Mosby, 1968:459.
- Pitts DG, Kay KR. The photo-ophthalmic threshold for the rabbit. *Am J Optom Arch Am Acad Optom* 1969;48:561-72.
- Pitts DG. A comparative study of the effects of ultraviolet radiation on the eye. *Am J Optom Arch Am Acad Optom* 1970;47:535-46.
- Pitts DG. The ocular ultraviolet action spectrum and protection criteria. *Health Phys* 1973;25:559-66.
- Pitts DG. The human ultraviolet action spectrum. *Am J Optom Arch Am Acad Optom* 1974;51:948-60.
- Pitts DG, Cullen AP, Hacker PD, Parr WD. Ocular ultraviolet effects from 295 nm to 400 nm in the rabbit eye. [DHEW (NIOSH) publication; 77-175] Cincinnati: U.S. Dept of Health, Education and Welfare National Institute for Occupational Safety and Health Division of Biomedical and Behavioral Science, 1977.
- Zuclich JA, Connolly JS. Ocular damage induced by near-ultraviolet laser radiation. *Invest Ophthalmol Vis Sci* 1976;15:760-4.
- Pitts DG, Lattimore MR. Protection against UVR using the Vistakon UV-Bloc soft contact lens. *Int Contact Lens Clin* 1987;14:22-9.
- Lattimore MR. Effect of ultraviolet radiation on the metabolism of the corneal epithelium of the rabbit. 1987. PhD Dissertation. University of Houston.
- Pitts E. Measurements of ultraviolet solar radiation. *Astrophys J* 1932;75:185-221.
- Zuclich JA, Connolly JS. Ocular damage induced by near-ultraviolet induced corneal damage. *Invest Ophthalmol Vis Sci* 1976;15:760-4.
- Ebbens RW, Sears D. Ocular effects of a 325 nm ultraviolet laser. *Am J Optom Physiol Opt* 1975;52:216-23.
- Benjamin WJ, Hill RM. Human corneal oxygen demand: the closed-eye interval. *Albrecht von Graefes Arch Klin Exp Ophthalmol* 1986;224:291-4.
- Holden BA, Sulonen J, Vannas A, Sweeney DF, Efron N. Direct *in vivo* measurement of corneal epithelial metabolic activity using a polarographic oxygen sensor. *Ophthalm Res* 1985;17:168-73.
- Maughar TF, Hill RM. Aerobic responses of the cornea to alkali measured *in vivo*. *Invest Ophthalmol Vis Sci* 1983;24:582-5.
- Murphy TM. Nucleic acids: interaction with solar UV radiation. *Curr Top Radiat Res Q* 1975;10:199-228.
- Ramabhadran TV. Effects of near ultraviolet and violet radiations (313-405 nm) on DNA, RNA, and protein synthesis in *E. coli B/r*: implications for growth delay. *Photochem Photobiol* 1975;22:117-23.
- Grabner G, Brenner W. Unscheduled DNA repair in the human cornea following solar simulating radiation. *Acta Ophthalmol (Kbh)* 1981;59:847-53.
- McLaren AD, Luse RA. Mechanism of inactivation of enzyme proteins by ultraviolet light. *Science* 1961;134:836-8.
- Augenstein L, Riley P. The inactivation of enzymes by ultraviolet light, 4: the nature and involvement of cystine disruption. *Photochem Photobiol* 1964;3:353-67.
- Vladimirov YA, Roschupkin DI, Fesenko EE. Photochemical reactions in amino acid residues and inactivation of enzymes during ultraviolet irradiation. *Photochem Photobiol* 1970;11:227-46.
- Grossweiner LJ. Photochemical inactivation of enzymes. *Curr Top Radiat Res Q* 1976;11:141-99.
- Grossweiner LJ. Photochemistry of proteins: a review. *Curr Eye Res* 1984;3:137-44.
- Kashket ER, Brodie AF. Oxidative phosphorylation in fractionated bacterial systems. 10: different roles for natural quinones of *Escherichia coli* w in oxidative metabolism. *J Biol Chem* 1963;238:2564-70.
- Ninneman H, Butler WL, Epel BL. Inhibition of respiration and destruction of cytochrome A3 by light in mitochondria and cytochrome oxidase from beef heart. *Biochem Biophys Acta* 1970;205:507-12.
- Ninneman H. Photoinhibition of isolated complexes I, II, and III of beef heart mitochondria. *FEBS Lett* 1974;39:353-8.
- Werben H, Lakshana BD, Jagger J. Near ultraviolet modification of *E. coli B. ubiquinone in vivo and in vitro*. *Photochem Photobiol* 1974;19:321-8.
- Crockett RS, Lawwill T. Oxygen dependence of damage by 435 nm light in cultured retinal epithelium. *Curr Eye Res* 1984;3:209-15.
- Griffin AC. Methoxsalen in ultraviolet carcinogenesis in the mouse. *J Invest Dermatol* 1959;32:367-72.
- Cloud TM, Hakim R, Griffin AC. Photosensitization of the eye with methoxsalen. I. Acute effects. *Arch Ophthalmol* 1960;64:346-51.
- Cloud TM, Hakim R, Griffin AC. Photosensitization of the eye with methoxsalen. II. Chronic effects. *Arch Ophthalmol* 1961;66:689-94.

32. Freeman RG, Troll D. Photosensitization of the eye by 8-methoxypsoralen. *J Invest Dermatol* 1969;53:449-53.
33. Parrish JA, Anderson RR, Urbach R, Pitts DG. UV-A Biological Effects of Radiation with Emphasis on Human Responses to Longwave Ultraviolet Radiation. New York: Plenum Press, 1978.
34. Pitts DG, Cullen AP. Ocular ultraviolet effects from 295 nm to 335 nm in the rabbit eye. [DHEW (NIOSH) publication; 77-130] Cincinnati: U.S. Dept of Health, Education and Welfare National Institute for Occupational Safety and Health Division of Biomedical and Behavioral Science, 1976.
35. Lerman S. Radiant energy and the eye. Functional Ophthalmology Series, vol 1. New York: MacMillan, 1980.
36. Kahn HA, Leibowitz HM, Ganley JP, Kini MM, Colton T, Wickerson RS, Dawabir TR. The Framingham Eye Study. I. Outline and major prevalence findings. *Am J Epidemiol* 1977;106:17-32. II. Association of ophthalmic pathology with single variables previously measured in the Framingham Heart Study. *Am J Epidemiol* 1977;106:33-41.
37. Hiller R, Giacometti L, Yuen K. Sunlight and cataract: an epidemiologic investigation. *Am J Epidemiol* 1977;105:450-9.
38. Taylor HR. The environment and the lens. *Br J Ophthalmol* 1980;64:303-10.
39. Hollows F, Moran D. Cataract—the ultraviolet risk factor. *Lancet* 1981;2:1249-51.
40. Brilliant LB, Grasset NC, Pokhrel RP, Kolstad A, Lepkowski JM, Brilliant GE, Hawks WN, Pararajasegaram R. Associations among cataract prevalence, sunlight hours, and altitude in the Himalayas. *Am J Epidemiol* 1983;118:250-64.
41. Hiller R, Sperduto RD, Ederer F. Epidemiologic associations with cataract in the 1971-1972 National Health and Nutrition Examination Survey. *Am J Epidemiol* 1983;118:239-49.
42. Leske MC, Sperduto RD. The epidemiology of senile cataracts: a review. *Am J Epidemiol* 1983;118:152-65.
43. Zigman S, Detiles M, Torczynski E. Sunlight and human cataracts. *Invest Ophthalmol Vis Sci* 1979;18:462-7.
44. Schmidt RE, Zuclich JA. Retinal lesions due to ultraviolet laser exposure. *Invest Ophthalmol Vis Sci* 1980;19:1166-75.
45. Ham WT, Ruffolo JJ, Mueller HA, Guerry D, Guerry RK. Action spectrum for retinal injury from near-ultraviolet radiation in the aphakic monkey. *Am J Ophthalmol* 1982;93:299-306.
46. Rapp LM, Jose JG, Pitts DG. DNA repair synthesis in the rat retina following *in vivo* exposures to 300 nm radiation. *Invest Ophthalmol Vis Sci* 1985;26:384-8.
47. Henton WW, Svkes SM. Recovery of absolute threshold with UV-A induced retinal damage. *Physiol Behav* 1984;32:949-54.
48. Waxler M. Long-term visual health risks from solar ultraviolet radiation. *Ophthalm Res* 1988;20:179-82.
49. Noel WK, Walker VS, Kang BS, Berman S. Retinal damage by light in rats. *Invest Ophthalmol Vis Sci* 1966;5:450-73.
50. Harwerth RS, Sperling HG. Prolonged color blindness induced by intense spectral lights in rhesus monkeys. *Science* 1971;174:520-3.
51. Lawwill T. Effects of prolonged exposure of rabbit retina to low intensity light. *Invest Ophthalmol Vis Sci* 1973;12:45-51.
52. Ham WT, Mueller HA, Sliney DH. Retinal sensitivity to damage from short wavelength light. *Nature* 1976;260:153-5.
53. Lawwill T, Crockett S, Currier G. Retinal damage secondary to chronic light exposure: thresholds and mechanisms. *Doc Ophthalmol* 1977;44:379-402.
54. Moon ME, Clarke AM, Ruffolo JJ, Mueller HA, Ham WT. Visual performance in the rhesus monkey after exposure to blue light. *Vision Res* 1978;18:1573-7.
55. Ham WT, Mueller HA, Ruffolo JJ, Clark AM. Sensitivity of the retina to radiation damage as a function of wavelength. *Photochem Photobiol* 1979;29:735-43.
56. Ham WT, Mueller HA, Ruffolo JJ. Retinal effects of blue light exposure. In: *Ocular Effects of Non-ionizing Radiation* [Proc Soc, Photo-Opt, Instrum, Eng. (USA): vol 229]. Washington: SPIE, 1980:46-50.
57. Ham WT, Ruffolo JJ, Mueller HA, Guerry D. The nature of retinal radiation damage: dependence on wavelength, power level, and exposure time. *Vision Res* 1980;20:1105-11.
58. Pitts DG, Bergmanson JPG, Chu LW-F. Rabbit eye exposure to broad-spectrum fluorescent light. *Acta Ophthalmol (Suppl) (Kbh)* 1983;159:1-54.
59. Zuclich JA, Kurtin WE. Oxygen dependence of near ultraviolet induced corneal damage. *Photochem Photobiol* 1977;25:133-5.
60. Zuclich JA. Ultraviolet induced damage in the primate cornea and retina. *Curr Eye Res* 1984;3:27-34.
61. Liang RA, Fischberg J, Chance B. Noninvasive measurements of pyridine nucleotide fluorescence from the cornea. *Invest Ophthalmol Vis Sci* 1980;19:96-102.
62. Masters BR. Noninvasive redox fluorometry: how light can be used to monitor alterations of corneal mitochondrial function. *Curr Eye Res* 1984;3:23-6.
63. Masters BR. Noninvasive corneal redox fluorometry. *Curr Top Eye Res* 1984;4:139-200.
64. Tsubota K, Laing RA, Kenyon KR. Noninvasive measurements of pyridine nucleotide and flavoprotein in the lens. *Invest Ophthalmol Vis Sci* 1987;28:785-9.

AUTHOR'S ADDRESS:

Morris R. Lattimore
U.S. Army Aeromedical Research Laboratory
P.O. Box 577
Fort Rucker, Alabama 36362-5292

Initial distribution

Commander
U.S. Army Natick Research
and Development Center
ATTN: Documents Librarian
Natick, MA 01760

Naval Submarine Medical
Research Laboratory
Medical Library, Naval Sub Base
Box 900
Groton, CT 05340

Commander/Director
U.S. Army Combat Surveillance
& Target Acquisition Lab
ATTN: DELCS-D
Fort Monmouth, NJ 07703-5304

Commander
10th Medical Laboratory
ATTN: Audiologist
APO NEW YORK 09180

Commander
Naval Air Development Center
Biophysics Lab
ATTN: G. Kydd
Code 60B1
Warminster, PA 18974

Naval Air Development Center
Technical Information Division
Technical Support Detachment
Warminster, PA 18974

Commanding Officer
Naval Medical Research
and Development Command
National Naval Medical Center
Bethesda, MD 20014

Under Secretary of Defense
for Research and Engineering
ATTN: Military Assistant
for Medical and Life Sciences
Washington, DC 20301

Commander
U.S. Army Research Institute
of Environmental Medicine
Natick, MA 01760

U.S. Army Avionics Research
and Development Activity
ATTN: SAVAA-P-TP
Fort Monmouth, NJ 07703-5401

U.S. Army Research and Development
Support Activity
Fort Monmouth, NJ 07703

Chief, Benet Weapons Laboratory
LCWSL, USA ARRADCOM
ATTN: DRDAR-LCB-TL
Watervliet Arsenal, NY 12189

Commander
Man-Machine Integration System
Code 602
Naval Air Development Center
Warminster, PA 18974

Commander
Naval Air Development Center
ATTN: Code 6021 (Mr. Brindle)
Warminster, PA 18974

Commanding Officer
Harry G. Armstrong Aerospace
Medical Research Laboratory
Wright-Patterson
Air Force Base, OH 45433

Director
Army Audiology and Speech Center
Walter Reed Army Medical Center
Washington, DC 20307-5001

Director
Walter Reed Army Institute
of Research
Washington, DC 20307-5100

HQ DA (DASG-PSP-0)
5109 Leesburg Pike
Falls Church, VA 22041-3258

Naval Research
Laboratory Library
Code 1433
Washington, DC 20375

Harry Diamond Laboratories
ATTN: Technical Information Branch
2800 Powder Mill Road
Adelphi, MD 20783-1197

U.S. Army Materiel Systems
Analysis Agency
ATTN: Reports Processing
Aberdeen Proving Ground
MD 21005-5017

U.S. Army Ordnance Center
and School Library
Building 3071
Aberdeen Proving Ground,
MD 21005-5201

U.S. Army Environmental Hygiene
Agency
Building E2100
Aberdeen Proving Ground,
MD 21010

Technical Library
Chemical Research
and Development Center
Aberdeen Proving Ground,
MD 21010-5423

Commander
U.S. Army Institute
of Dental Research
Walter Reed Army Medical Center
Washington, DC 20307-5300

Naval Air Systems Command
Technical Air Library 950D
Rm 278, Jefferson Plaza II
Department of the Navy
Washington, DC 20361

Naval Research Laboratory Library
Shock and Vibration Information Center, Code 5804
Washington, DC 20375

Director
U.S. Army Human Engineering Laboratory
ATTN: Technical Library
Aberdeen Proving Ground,
MD 21005-5001

Commander
U.S. Army Test
and Evaluation Command
ATTN: AMSTE-AD-H
Aberdeen Proving Ground,
MD 21005-5055

Director
U.S. Army Ballistic
Research Laboratory
ATTN: DRXBR-OD-ST Tech Reports
Aberdeen Proving Ground,
MD 21005-5066

Commander
U.S. Army Medical Research
Institute of Chemical Defense
ATTN: SGRD-UV-AO
Aberdeen Proving Ground,
MD 21010-5425

Commander
U.S. Army Medical Research
and Development Command
ATTN: SGRD-RMS (Ms. Madigan)
Fort Detrick, Frederick,
MD 21701

Commander
U.S. Army Medical Research
Institute of Infectious Diseases
Fort Detrick, Frederick,
MD 21701

Director, Biological
Sciences Division
Office of Naval Research
600 North Quincy Street
Arlington, VA 22217

Commander
U.S. Army Materiel Command
ATTN: AMCDE-XS
5001 Eisenhower Avenue
Alexandria, VA 22333

Commandant
U.S. Army Aviation
Logistics School
ATTN: ATSQ-TDN
Fort Eustis, VA 23604

U.S. Army Training
and Doctrine Command
ATTN: ATCD-ZX
Fort Monroe, VA 23651

Structures Laboratory Library
USARTL-AVSCOM
NASA Langley Research Center
Mail Stop 266
Hampton, VA 23665

Naval Aerospace Medical
Institute Library
Bldg 1953, Code 102
Pensacola, FL 32508

Command Surgeon
U.S. Central Command
MacDill Air Force Base
FL 33608

Air University Library
(AUL/LSE)
Maxwell AFB, AL 36112

Commander
U.S. Army Biomedical Research
and Development Laboratory
ATTN: SGRD-UBZ-I
Fort Detrick, Frederick,
MD 21701

Defense Technical
Information Center
Cameron Station
Alexandria, VA 22313

U.S. Army Foreign Science
and Technology Center
ATTN: MTZ
220 7th Street, NE
Charlottesville, VA 22901-5396

Director,
Applied Technology Laboratory
USARTL-AVSCOM
ATTN: Library, Building 401
Fort Eustis, VA 23604

U.S. Army Training
and Doctrine Command
ATTN: Surgeon
Fort Monroe, VA 23651-5000

Aviation Medicine Clinic
TMC #22, SAAF
Fort Bragg, NC 28305

U.S. Air Force Armament
Development and Test Center
Eglin Air Force Base, FL 32542

U.S. Army Missile Command
Redstone Scientific
Information Center
ATTN: Documents Section
Redstone Arsenal, AL 35898-5241

U.S. Army Research and Technology
Laboratories (AVSCOM)
Propulsion Laboratory MS 302-2
NASA Lewis Research Center
Cleveland, OH 44135

AFAMRL/HEX
Wright-Patterson AFB, OH 45433

University of Michigan
NASA Center of Excellence
in Man-Systems Research
ATTN: R. G. Snyder, Director
Ann Arbor, MI 48109

John A. Dellinger,
Southwest Research Institute
P. O. Box 28510
San Antonio, TX 78284

Product Manager
Aviation Life Support Equipment
ATTN: AMCPM-ALSE
4300 Goodfellow Blvd.
St. Louis, MO 63120-1798

Commander
U.S. Army Aviation
Systems Command
ATTN: AMSAV-ED
4300 Goodfellow Blvd
St. Louis, MO 63120

Commanding Officer
Naval Biodynamics Laboratory
P.O. Box 24907
New Orleans, LA 70189

U.S. Army Field Artillery School
ATTN: Library
Snow Hall, Room 14
Fort Sill, OK 73503

Commander
U.S. Army Health Services Command
ATTN: HSOP-SO
Fort Sam Houston, TX 78234-6000

U.S. Air Force Institute
of Technology (AFIT/LDEE)
Building 640, Area B
Wright-Patterson AFB, OH 45433

Henry L. Taylor
Director, Institute of Aviation
University of Illinois-
Willard Airport
Savoy, IL 61874

COL Craig L. Urbauer, Chief
Office of Army Surgeon General
National Guard Bureau
Washington, DC 50310-2500

Commander
U.S. Army Aviation
Systems Command
ATTN: SGRD-UAX-AL (MAJ Lacy)
4300 Goodfellow Blvd., Bldg 105
St. Louis, MO 63120

U.S. Army Aviation Systems Command
Library and Information
Center Branch
ATTN: AMSAV-DIL
4300 Goodfellow Blvd
St. Louis, MO 63120

Federal Aviation Administration
Civil Aeromedical Institute
CAMI Library AAC 64D1
P.O. Box 25082
Oklahoma City, OK 73125

Commander
U.S. Army Academy
of Health Sciences
ATTN: Library
Fort Sam Houston, TX 78234

Commander
U.S. Army Institute
of Surgical Research
ATTN: SGRD-USM (Jan Duke)
Fort Sam Houston, TX 78234-6200

Director of Professional Services
AFMSC/GSP
Brooks Air Force Base, TX 78235

U.S. Army Dugway Proving Ground
Technical Library
3Bldg 5330
Dugway, UT 84022

U.S. Army Yuma Proving Ground
Technical Library
Yuma, AZ 85364

AFFTC Technical Library
6520 TESTG/ENXL
Edwards Air Force Base,
CAL 93523-5000

Commander
Code 3431
Naval Weapons Center
China Lake, CA 93555

Aeromechanics Laboratory
U.S. Army Research
and Technical Labs
Ames Research Center,
M/S 215-1
Moffett Field, CA 94035

Sixth U.S. Army
ATTN: SMA
Presidio of San Francisco,
CA 94129

Commander
U.S. Army Aeromedical Center
Fort Rucker, AL 36362

Commander, U.S. Army
Aviation Center
Directorate
of Combat Developments
Bldg 507

U.S. Air Force School
of Aerospace Medicine
Strughold Aeromedical Library
Documents Section, USAFSAM/TSK-4
Brooks Air Force Base, TX 78235

Dr. Diane Damos
Department of Human Factors
ISSM, USC
Los Angeles, CA 90089-0021

U.S. Army White Sands
Missile Range
Technical Library Division
White Sands Missile Range,
NM 88002

U.S. Army Aviation Engineering
Flight Activity
ATTN: SAVTE-M (Tech Lib)
Stop 217
Edwards Air Force Base,
CA 93523-5000

Ms. Sandra G. Hart
Ames Research Center
MS 239-5
Moffett Field, CA 94035

Commander
Letterman Army Institute
of Research
ATTN: Medical Research Library
Presidio of San Francisco,
CA 94129

Director
Naval Biosciences Laboratory
Naval Supply Center, Bldg 844
Oakland, CA 94625

Commander
U.S. Army Medical Materiel
Development Activity
Fort Detrick, Frederick,
MD 21701-5009

Directorate
of Training Development
Bldg 502

Fort Rucker, AL 36362

Chief
Army Research Institute
Field Unit
Fort Rucker, AL 36362

Commander
U.S. Army Safety Center
Fort Rucker, AL 36362

U.S. Army Aircraft Development
Test Activity
ATTN: STEBG-MP-QA
Cairns AAF
Fort Rucker, AL 36362

Commander
U.S. Army Medical Research
and Development Command
ATTN: SGRD-PLC (COL Sedge)
Fort Detrick, Frederick
MD 21701

Fort Rucker, AL 36362

Chief
Human Engineering Laboratory
Field Unit
Fort Rucker, AL 36362

Commander
U.S. Army Aviation Center
and Fort Rucker
ATTN: ATZQ-T-ATL
Fort Rucker, AL 36362

President
U.S. Army Aviation Board
Cairns AAF
Fort Rucker, AL 36362